



# Characterization of a novel nonpeptide vasopressin V<sub>2</sub>-agonist, OPC-51803, in cells transfected human vasopressin receptor subtypes

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**1** We discovered the first nonpeptide arginine-vasopressin (AVP) V<sub>2</sub>-receptor agonist, OPC-51803. Pharmacological properties of OPC-51803 were elucidated using HeLa cells expressing human AVP receptor subtypes (V<sub>2</sub>, V<sub>1a</sub> and V<sub>1b</sub>) and compared with those of 1-desamino-8-D-arginine vasopressin (dDAVP), a peptide V<sub>2</sub>-receptor agonist.

**2** OPC-51803 and dDAVP displaced [<sup>3</sup>H]-AVP binding to human V<sub>2</sub>- and V<sub>1a</sub>-receptors with *K<sub>i</sub>* values of 91.9 ± 10.8 nM (*n* = 6) and 3.12 ± 0.38 nM (*n* = 6) for V<sub>2</sub>-receptors, and 819 ± 39 nM (*n* = 6) and 41.5 ± 9.9 nM (*n* = 6) for V<sub>1a</sub>-receptors, indicating that OPC-51803 was about nine times more selective for V<sub>2</sub>-receptors, similar to the selectivity of dDAVP. OPC-51803 scarcely displaced [<sup>3</sup>H]-AVP binding to human V<sub>1b</sub>-receptors even at 10<sup>−4</sup> M, while dDAVP showed potent affinity to human V<sub>1b</sub>-receptors with the *K<sub>i</sub>* value of 13.7 ± 3.2 nM (*n* = 4).

**3** OPC-51803 concentration-dependently increased cyclic adenosine 3', 5'-monophosphate (cyclic AMP) production in HeLa cells expressing human V<sub>2</sub>-receptors with an EC<sub>50</sub> value of 189 ± 14 nM (*n* = 6). The concentration-response curve for cyclic AMP production induced by OPC-51803 was shifted to the right in the presence of a V<sub>2</sub>-antagonist, OPC-31260.

**4** At 10<sup>−5</sup> M, OPC-51803 did not increase the intracellular Ca<sup>2+</sup> concentration ([Ca<sup>2+</sup>]<sub>i</sub>) in HeLa cells expressing human V<sub>1a</sub>-receptors. On the other hand, dDAVP increased [Ca<sup>2+</sup>]<sub>i</sub> in HeLa cells expressing human V<sub>1a</sub>- and V<sub>1b</sub>-receptors in a concentration-dependent fashion.

**5** From these results, OPC-51803 has been confirmed to be the first nonpeptide agonist for human AVP V<sub>2</sub>-receptors without agonistic activities for V<sub>1a</sub>- and V<sub>1b</sub>-receptors. OPC-51803 may be useful for the treatment of AVP-deficient pathophysiological states and as a tool for AVP researches.

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**Abbreviations:** ACTH, adrenocorticotrophic hormone; AVP, arginine vasopressin; BSA, bovine serum albumin; EC<sub>50</sub>, concentration exerting 50% effectiveness of maximal response; IC<sub>50</sub>, concentration exerting 50% inhibition; cyclic AMP, cyclic adenosine 3', 5'-monophosphate; dDAVP, 1-desamino-8-D-arginine vasopressin; DMSO, dimethylsulphoxide; *K<sub>D</sub>*, dissociation constant; DMEM, Dulbecco's minimum Eagle's medium; EGTA, ethylene glycol-bis (β-aminoethylether)-*N,N,N',N'*-tetraacetic acid; FBS, foetal bovine serum; Fura-2-AM, fura-2 acetoxymethyl ester; G418, geneticin disulphate; HEPES, 2-[4-(2-hydroxyethyl)-1-piperazinyl] ethanesulphonic acid; *K<sub>i</sub>*, inhibition constant; [Ca<sup>2+</sup>]<sub>i</sub>, intracellular Ca<sup>2+</sup> concentration; IBMX, 3-isobutyl-1-methylxanthine; B<sub>max</sub>, number of binding sites; PBS, phosphate-buffered saline; Triton X-100, polyethylene glycol mono-*p*-isooctylphenyl ether; SDS, sodium dodecyl sulphate

## Introduction

Vasopressin (AVP) is a neurohypophysial hormone which is synthesized in the hypothalamus and released from the pituitary. AVP plays an important role in the regulation of body fluid homeostasis. Physiological actions of AVP are exerted through seven transmembrane G-protein-coupled receptors (Laszlo *et al.*, 1991; Liu & Wess, 1996). V<sub>1a</sub>-receptors

are the most widespread and are coupled with G<sub>q/11</sub>; they are responsible for vasoconstriction, platelet aggregation, glycolysis and so on, and V<sub>1b</sub>-receptors are also coupled with G<sub>q/11</sub> and are responsible for adrenocorticotrophic hormone (ACTH) secretion, *via* stimulation of inositol phosphate turnover. V<sub>2</sub>-receptors are coupled to G<sub>s</sub>, and responsible for promotion of water reabsorption in the collecting ducts of the kidney *via* stimulation of cyclic adenosine 3', 5'-monophosphate (cyclic AMP) production.

In the 1990's, we reported orally effective nonpeptide antagonists of AVP receptors, V<sub>1a</sub>-selective OPC-21268 (Ogawa *et al.*, 1993; Yamamura *et al.*, 1991), V<sub>2</sub>-selective OPC-31260 (Ogawa *et al.*, 1996; Yamamura *et al.*, 1992) and OPC-41061 (Kondo *et al.*, 1999; Yamamura *et al.*, 1998). Then

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several nonpeptide antagonists of AVP receptors, such as V<sub>1a</sub>-selective SR-49059 (Serradeil-Le Gal *et al.*, 1993), V<sub>2</sub>-selective SR-121463A (Serradeil-Le Gal *et al.*, 1996), and VPA-985 (Albright *et al.*, 1998), and nonselective YM-087 (Tahara *et al.*, 1997) followed. These nonpeptide antagonists could overcome the problem of intrinsic agonistic activity associated with peptide vasopressin antagonists. Since peptide antagonists have an intrinsic agonistic activity, they acted as an agonist when tested in human. Though it has been difficult to develop antagonists without an agonistic activity for human AVP V<sub>2</sub>-receptors by peptide analogues, nonpeptide compounds succeeded in acting as an antagonist in human (Ohnishi *et al.*, 1993; 1995). The approach for developing the nonpeptide ligands for peptide hormone receptors may enable us to produce functionally separate ligands, agonist and antagonist, and elucidate the mechanisms of receptor-ligand interactions.

Recently, nonpeptide agonists have been targeted for the seven transmembrane G-protein-coupled receptors as well as antagonists. In fact, nonpeptide agonists have been reported in the angiotensin system (Kivlighn *et al.*, 1995; Perlman *et al.*, 1995) and the cholecystokinin system (Aquino *et al.*, 1996; Willson *et al.*, 1996). During our development of nonpeptide antagonists for AVP receptors, some compounds were also found to possess antidiuretic effects in rats. Since molecular cloning of human AVP receptor subtypes was identified (Birnbauer *et al.*, 1992; de Keyser *et al.*, 1994; Hirasawa *et al.*, 1994; Lolait *et al.*, 1992; Sugimoto *et al.*, 1994; Thibonnier *et al.*, 1994), we designed the stable expression of each human AVP receptor in HeLa cells (Yamamura *et al.*, 1998) and attempted to develop functionally acting, human V<sub>2</sub>-receptor agonists. After the lead optimization of congeners of OPC-31260 and OPC-41061, OPC-51803 was selected as the first nonpeptide AVP V<sub>2</sub>-receptor agonist. In this study, we used cells stably expressing each human AVP receptor subtype (V<sub>2</sub>, V<sub>1a</sub>, and V<sub>1b</sub>) and tried to characterize the binding and functional properties of OPC-51803, (5*R*)-2-[1-(2-chloro-4-(1-pyrrolidinyl) benzoyl)-2,3,4,5-tetrahydro-1*H*-1-benzazepin-5-yl]-*N*-isopropylacetamide (Figure 1) and compared it with dDAVP, a standard peptide AVP V<sub>2</sub>-receptor agonist.

## Methods

### Materials

OPC-51803 and OPC-31260 were synthesized by the Tokushima Second Factory, Otsuka Pharmaceutical Co., Ltd. (Tokyo, Japan). AVP was purchased from Peptide Institute Inc. (Osaka, Japan), [<sup>3</sup>H]-AVP from NEN<sup>TM</sup> Lifescience Products, Inc. (Boston, MA, U.S.A.), dDAVP, [deamino-Pen<sup>1</sup>, O-Me-Tyr<sup>2</sup>, Arg<sup>8</sup>]-vasopressin, penicillin, streptomycin, 3-isobutyl-1-methylxanthine (IBMX), ethylene glycol-bis (β-aminoethylether)-*N,N,N',N'*-tetraacetic acid (EGTA), bovine serum albumin (BSA), MnCl<sub>2</sub>, and trypsin inhibitor from Sigma (St. Louis, MO, U.S.A.), lipofectamine, foetal bovine serum (FBS), and trypsin-EDTA from GIBCO BRL (Rockville, MD, U.S.A.), Dulbecco's minimum Eagle's medium (DMEM) from Nissui Pharmaceutical Co. (Tokyo, Japan), phosphate-buffered salts from Takara Shuzo Co., Ltd. (Kusatsu, Japan), 2-[4-(2-hydroxyethyl)-1-piperazinyl]ethanesulphonic acid (HEPES), Fura-2 and Fura-2 acetoxymethyl ester (Fura-2-AM) from Dojindo Laboratories (Kumamoto, Japan), polyethylene glycol mono-*p*-isooctylphenyl ether (Triton X-100) from Nacalai Tesque (Kyoto, Japan), and

geneticin disulphate (G418) and other chemicals from Wako Pure Chemicals (Osaka, Japan).

### Preparation of HeLa cells expressing human AVP V<sub>2</sub>, V<sub>1a</sub>, and V<sub>1b</sub>-receptors

Cloned cDNAs for AVP receptors were subcloned into pBluescript KS II(+) (Stratagene, La Jolla, CA, U.S.A.). Nucleotide sequence analysis was performed using an ABI 373A DNA Sequencer (Applied Biosystems, Inc., Foster City, CA, U.S.A.) and the obtained clones were confirmed to be identical to the previously reported human AVP-receptor cDNAs. These cDNA fragments were ligated in SRα promoter-based mammalian expression vectors psvk3 or pME18S and resulting constructs were used for transfection.

HeLa cells, human endocervical carcinoma cell line, grown in 5% CO<sub>2</sub> at 37°C, were plated at a density of 1 × 10<sup>5</sup> cells in 10-cm plate with 10 ml of DMEM supplemented containing 10% FBS and penicillin/streptomycin, and allowed to attach overnight. Each expression vector for vasopressin receptors and pSV2neo vector (Clontech, Palo Alto, CA, U.S.A.), which contained a neomycin resistance gene for selection of stable transformants, were co-transfected into HeLa cells by the lipofectamine method (Hawley-Nelson *et al.*, 1993). After overnight transfection, cells were fed with fresh medium, allowed to grow for 2–3 days, adjusted to a density of 1 × 10<sup>3</sup> cells per plate, and incubated for an additional 24 h. Cells were continually selected for 3–4 weeks in a medium containing 400–800 µg ml<sup>-1</sup> of the antibiotic G418. Single colonies were then isolated, expanded, and harvested for radioligand binding assays to measure the expression of receptors. Cells were maintained in the medium containing 200 µg ml<sup>-1</sup> of G418. The medium was changed every 3 days, and the cells were subcultured after trypsinization.

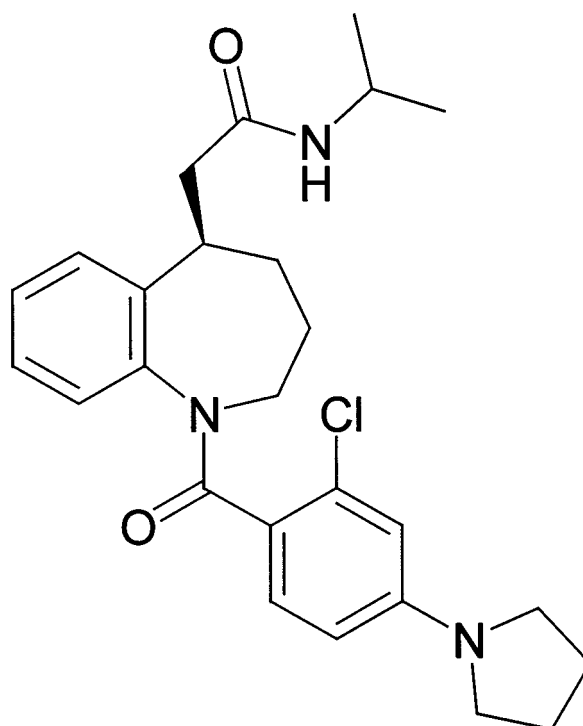


Figure 1 Structure of OPC-51803.

### Radioligand binding assay to HeLa cells expressing human AVP receptors

The experiments were performed after each cell reached confluence in 12-well (V<sub>1a</sub> and V<sub>1b</sub>) or 24-well (V<sub>2</sub>) dishes. After washing three times with ice-cold phosphate-buffered saline (PBS), the cells were incubated with [<sup>3</sup>H]-AVP with or without unlabelled AVP (final concentration of 1  $\mu$ M) to determine the total binding or the nonspecific binding, respectively in DMEM containing 0.3% BSA brought to a pH of 7.4 with 10 mM HEPES-NaOH. For the competition experiments, OPC-51803 dissolved in dimethylsulphoxide (DMSO), AVP and dDAVP dissolved in saline were diluted with DMEM medium and added to each well at several appropriate concentrations. After incubation at 4°C for 2 h, the reaction was stopped by removing the buffer, and the cells were rinsed three times with ice-cold PBS. The cells were lysed with 500  $\mu$ l (12-well) or 250  $\mu$ l (24-well) of 0.1 N NaOH containing 0.1% sodium dodecyl sulphate (SDS), transferred into scintillation vials, and mixed with 5 ml of Aquazol II (Packard, Tokyo, Japan). Radioactivity was detected using a liquid scintillation counter (LSC-1050, Aloka, Tokyo, Japan). The cells in some wells were lysed in 0.1 N NaOH and protein contents were determined by the dye method (Bradford, 1976).

### Cyclic AMP production in HeLa cells expressing V<sub>2</sub>-receptors

After reaching confluence in 24-well dishes, the cells were washed twice with ice-cold PBS and incubated with the DMEM containing 0.3% BSA and 1 mM IBMX brought to a pH of 7.4 with 10 mM HEPES-NaOH. After the addition of certain concentrations of OPC-51803, AVP, or dDAVP, the incubation was carried out at 37°C for 10 min. For the inhibition experiments, OPC-51803 was added following OPC-31260, a nonpeptide V<sub>2</sub> antagonist, at 10<sup>-7</sup> M and 10<sup>-6</sup> M. The reaction was stopped by aspiration of the medium and the cells were rinsed twice with ice-cold PBS. Cyclic AMP was extracted from the cells by adding 250  $\mu$ l of 0.1 N HCl and stored at -20°C until cyclic AMP determination using a radioimmunoassay kit (Yamasa, Tokyo, Japan).

### Intracellular Ca<sup>2+</sup> concentration in HeLa cells expressing V<sub>1a</sub>- or V<sub>1b</sub>-receptors

The harvested cells were suspended in Ca<sup>2+</sup>-loading buffer (in mM: NaCl 140, KCl 4, MgCl<sub>2</sub> 1, CaCl<sub>2</sub> 1.25, Na<sub>2</sub>HPO<sub>4</sub> 1, glucose 11, BSA 0.2%, and HEPES 5, brought to a pH of 7.4 with NaOH) and incubated at 37°C for 30 min with Fura-2-AM at 5  $\mu$ M under protection from light. After the incubation, the cells were washed twice and resuspended with Ca<sup>2+</sup>-loading buffer.

The fluorescence was monitored with fluorescence spectrophotometer (F-2000, Hitachi, Tokyo, Japan) by excitation at 340 and 380 nm and emission at 510 nm. Maximum and minimum fluorescence levels were measured using 0.1% Triton X-100 and 3 mM EGTA, respectively. Autofluorescence of the cells was measured by applying 5 mM MnCl<sub>2</sub>. Actual [Ca<sup>2+</sup>]<sub>i</sub> was calculated using the equation of Grynkiewicz *et al.* (1985) with a K<sub>D</sub> for Fura-2 of 224 nM. OPC-51803 dissolved in DMSO and AVP and dDAVP dissolved in PBS were applied cumulatively, and concentration-dependent curves were obtained. For the inhibition experiments, [deamino-Pen<sup>1</sup>, O-Me-Tyr<sup>2</sup>, Arg<sup>8</sup>]-vasopressin (a peptide V<sub>1</sub>-antagonist) at 10<sup>-7</sup> M and 10<sup>-6</sup> M dissolved in PBS, were pretreated with the cells

before adding test compounds. The test compounds were confirmed not to interfere with the Fura-2-Ca<sup>2+</sup> fluorescence at used concentrations.

### Data analysis

All data are expressed as the mean  $\pm$  s.e.mean. In saturation binding experiments, the dissociation constant (K<sub>D</sub>) and the number of binding sites (B<sub>max</sub>) were determined by Scatchard analysis (Scatchard, 1949). The IC<sub>50</sub> of OPC-51803, AVP, and dDAVP were determined by displacement curves of [<sup>3</sup>H]-AVP binding. The inhibition constant (K<sub>i</sub>) was calculated from the equation of K<sub>i</sub> = IC<sub>50</sub>/(1 + [L]/K<sub>D</sub>), where IC<sub>50</sub> is the concentration exerting 50% inhibition and [L] is the concentration of a radiolabelled ligand (Cheng & Prusoff, 1973). Regarding the activity of each compound on cyclic AMP production and [Ca<sup>2+</sup>]<sub>i</sub>, EC<sub>50</sub> was determined as that concentration which produces 50% of the maximal response.

## Results

### Binding affinities of OPC-51803, AVP, and dDAVP for human AVP receptors

HeLa cells expressing human AVP receptors (V<sub>2</sub>-, V<sub>1a</sub>-, or V<sub>1b</sub>-subtype) showed constant K<sub>D</sub> and B<sub>max</sub> throughout repeated cultures (V<sub>2</sub>: passage 9–18, V<sub>1a</sub>: passage 8–14, and V<sub>1b</sub>: passage 10–20). These K<sub>D</sub> and B<sub>max</sub> values are listed in Table 1. The stimulation by AVP increased cyclic AMP production in cells expressing V<sub>2</sub>-receptors and [Ca<sup>2+</sup>]<sub>i</sub> in cells expressing V<sub>1a</sub>- and V<sub>1b</sub>-receptors, indicating that these transfected HeLa cells acted functionally throughout the experiments.

Unlabelled AVP displaced [<sup>3</sup>H]-AVP binding for all three receptor subtypes with similar K<sub>i</sub> values (Figure 2). These K<sub>i</sub> values are also comparable to K<sub>D</sub> values of [<sup>3</sup>H]-AVP (Table 2). OPC-51803 displaced [<sup>3</sup>H]-AVP binding to human V<sub>2</sub>- and V<sub>1a</sub>-receptors in a concentration-dependent manner, although it scarcely displaced [<sup>3</sup>H]-AVP binding to human V<sub>1b</sub>-receptors even at 10<sup>-4</sup> M (Figure 2). The K<sub>i</sub> value of OPC-51803 for V<sub>2</sub>-receptors was 91.9  $\pm$  10.8 nM (n = 6), and that for V<sub>1a</sub>-receptors was 819  $\pm$  39 M (n = 6), which showed about nine times more selectivity for V<sub>2</sub>-receptors than V<sub>1a</sub>-receptors. While dDAVP is well-known as a selective V<sub>2</sub>-agonist, it displaced [<sup>3</sup>H]-AVP binding not only to V<sub>2</sub>-receptors but also to V<sub>1a</sub>- and V<sub>1b</sub>-receptors. These results are consistent with previous observations (Chini *et al.*, 1995; Pettibone *et al.*, 1992; Saito *et al.*, 1997). Receptor selectivity of dDAVP was about 13 times less potent for V<sub>1a</sub>-receptors and four times less potent for V<sub>1b</sub>-receptors than for V<sub>2</sub>-receptors.

**Table 1** Dissociation constants (K<sub>D</sub>) for AVP and numbers of AVP receptors (B<sub>max</sub>) in HeLa cells expressing human AVP V<sub>2</sub>-, V<sub>1a</sub>-, and V<sub>1b</sub>-receptors

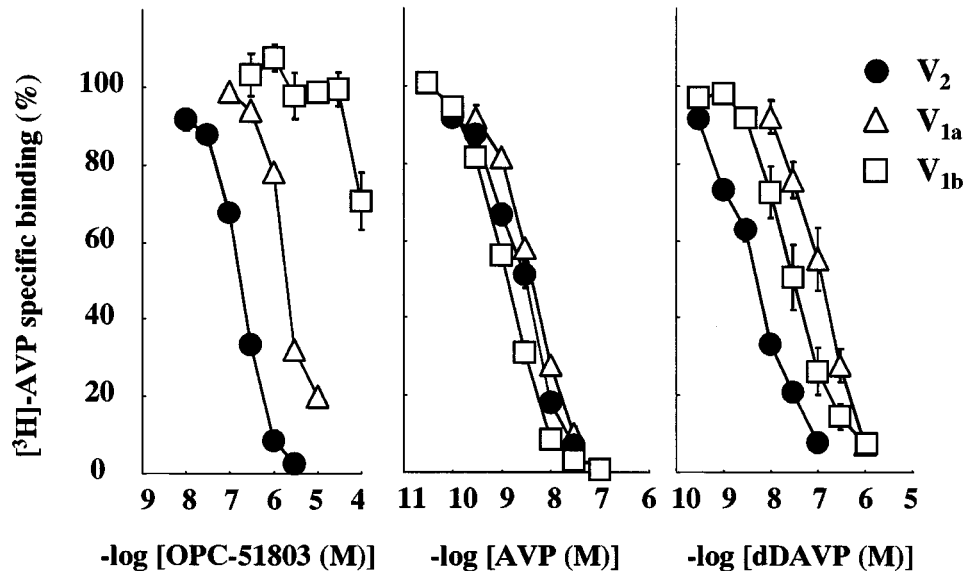
	HeLa cells transfected each human AVP receptor gene		
	V <sub>2</sub> -receptors	V <sub>1a</sub> -receptors	V <sub>1b</sub> -receptors
K <sub>D</sub> (nM)	3.22 $\pm$ 0.11	2.20 $\pm$ 0.09	2.43 $\pm$ 0.25
B <sub>max</sub> (fmol mg <sup>-1</sup> protein)	3354 $\pm$ 564	446 $\pm$ 63	98 $\pm$ 9

Values are expressed as the mean  $\pm$  s.e.mean of six or seven experiments performed in duplicate.

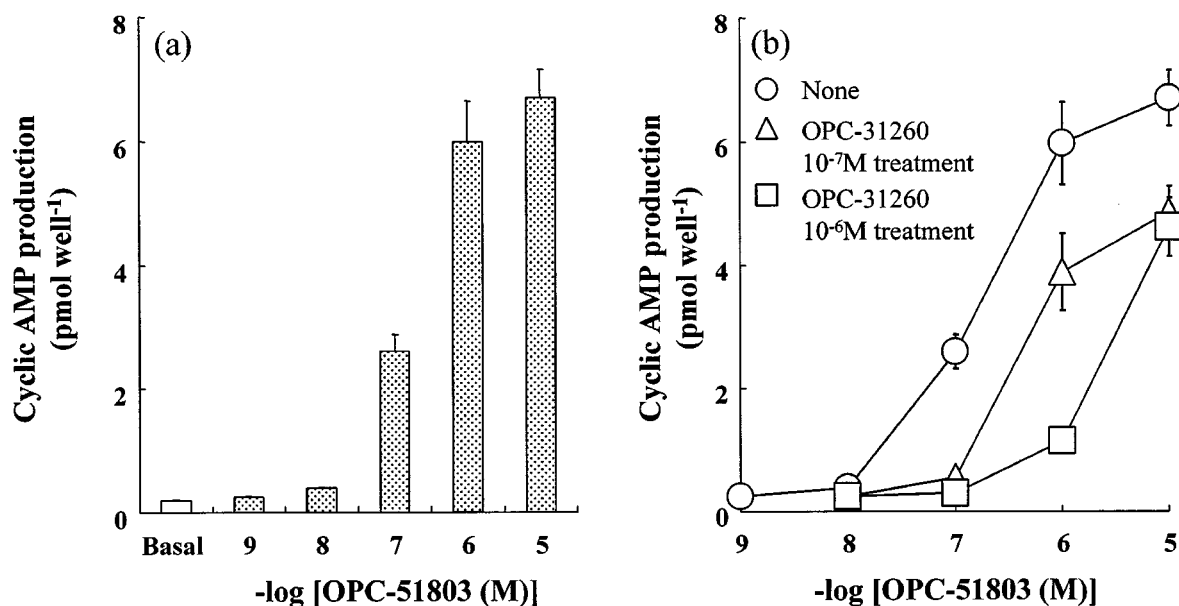
### Cyclic AMP production in HeLa cells expressing V<sub>2</sub>-receptors

To confirm the functional properties of OPC-51803, cyclic AMP production was examined using intact cells expressing V<sub>2</sub>-receptors. OPC-51803 concentration-dependently in-

creased cyclic AMP production in HeLa cells expressing human V<sub>2</sub>-receptors at OPC-51803 concentrations of 10<sup>-9</sup>–10<sup>-5</sup> M (Figure 3a). At 10<sup>-5</sup> M, cyclic AMP production of 6.70 ± 0.45 pmol well<sup>-1</sup> was seen and the production was 35 times higher than the basal value (0.19 ± 0.01 pmol well<sup>-1</sup>). The EC<sub>50</sub> value of OPC-51803 on cyclic AMP production



**Figure 2** Displacements of OPC-51803, AVP, and dDAVP on [<sup>3</sup>H]-AVP binding to human V<sub>2</sub>-, V<sub>1a</sub>-, and V<sub>1b</sub>-receptors in HeLa cells. Values are expressed as the mean ± s.e.mean of 4–6 experiments performed in duplicate.



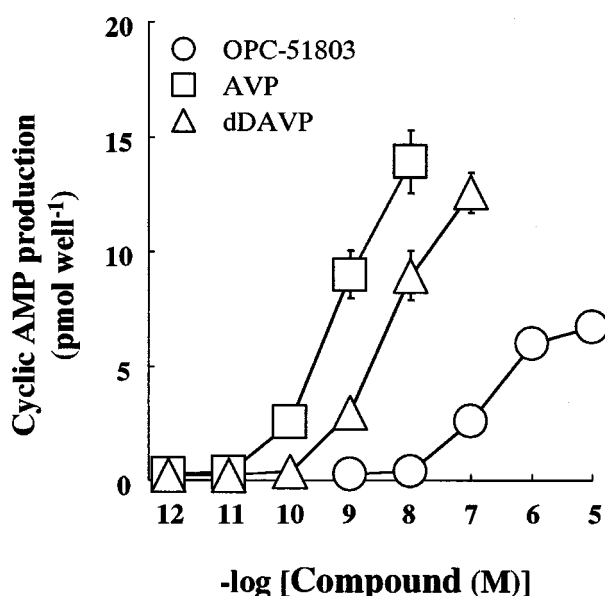
**Figure 3** (a) OPC-51803-induced increases in cyclic AMP production and (b) the inhibitory effect of OPC-31260 in HeLa cells expressing human V<sub>2</sub>-receptors. Values are expressed as the mean ± s.e.mean of five experiments performed in triplicate.

**Table 2** Inhibition constant (*K<sub>i</sub>*) and effective concentration (EC<sub>50</sub>) of OPC-51803, AVP, and dDAVP in human HeLa cells expressing V<sub>2</sub>-, V<sub>1a</sub>-, and V<sub>1b</sub>-receptors

	V <sub>2</sub> -receptors		V <sub>1a</sub> -receptors		V <sub>1b</sub> -receptors	
	K <sub>i</sub> (nM)	EC <sub>50</sub> (nM)	K <sub>i</sub> (nM)	EC <sub>50</sub> (nM)	K <sub>i</sub> (nM)	EC <sub>50</sub> (nM)
OPC-51803	91.9 ± 10.8	189 ± 14	819 ± 39	NE	NE	ND
AVP	1.44 ± 0.23	0.56 ± 0.10	1.51 ± 0.11	0.86 ± 0.17	0.46 ± 0.08	3.00 ± 0.50
dDAVP	3.12 ± 0.38	4.77 ± 1.43	41.5 ± 9.9	28.9 ± 2.6	13.7 ± 3.2	25.4 ± 2.5

Values are expressed as the mean ± s.e.mean of 4–6 experiments. NE; no effect, ND; not determined.

was  $189 \pm 14$  nM ( $n=5$ , Table 2). OPC-31260, a selective  $V_2$ -receptor antagonist, did not influence basal cyclic AMP production (data not shown). In the presence of OPC-31260 at  $10^{-7}$  M and  $10^{-6}$  M, the concentration-response curve of OPC-51803 was shifted to the right (Figure 3b), which suggests that the OPC-51803-induced increases in cyclic AMP are mediated through activation of  $V_2$ -receptors. AVP and dDAVP also enhanced cyclic AMP production in HeLa cells expressing human  $V_2$ -receptors with  $EC_{50}$  values of  $0.56 \pm 0.10$  nM and  $4.77 \pm 1.43$  nM ( $n=5$  each, Table 2)



**Figure 4** Comparison of OPC-51803 with AVP and dDAVP on cyclic AMP production in HeLa cells expressing human  $V_2$ -receptors. Values are expressed as the mean  $\pm$  s.e. mean of five experiments performed in triplicate.

and the maximal increases in cyclic AMP were higher than that by OPC-51803 (Figure 4).

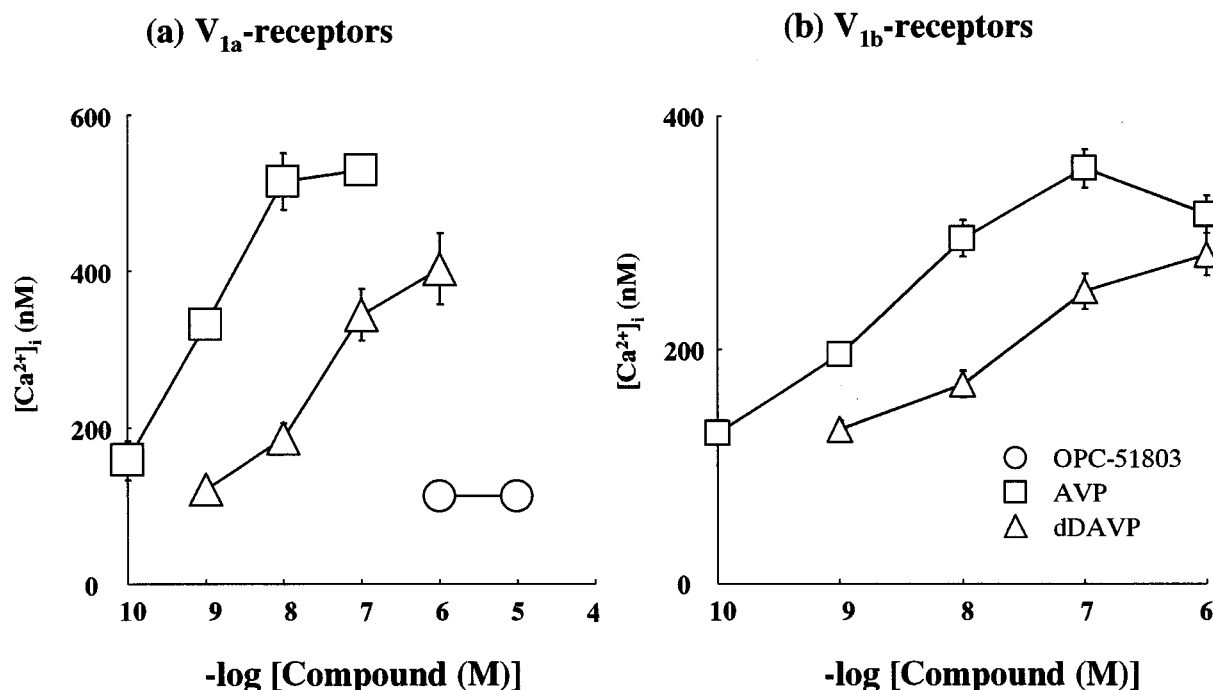
#### Intracellular $Ca^{2+}$ concentration in human HeLa cells expressing $V_{1a}$ - and $V_{1b}$ -receptors

We further examined agonistic action on human  $V_{1a}$ - and  $V_{1b}$ -receptors by measuring the change in  $[Ca^{2+}]_i$ , using the Fura-2 fluorescence method (Figure 5 and Table 2). AVP at concentrations of  $10^{-10}$ – $10^{-7}$  M increased  $[Ca^{2+}]_i$  from  $104 \pm 9$  to  $530 \pm 15$  nM in HeLa cells expressing  $V_{1a}$ -receptors ( $n=5$ ). dDAVP at  $10^{-9}$ – $10^{-6}$  M also raised  $[Ca^{2+}]_i$  from  $115 \pm 12$  to  $403 \pm 45$  nM in these cells ( $n=5$ ). Concentration-response curves of AVP and dDAVP for  $[Ca^{2+}]_i$  were shifted to the right in the presence of a peptide  $V_{1a}$ -antagonist at  $10^{-7}$  and  $10^{-6}$  M (data not shown). On the other hand, OPC-51803 at  $10^{-6}$  and  $10^{-5}$  M did not increase  $[Ca^{2+}]_i$  in  $V_{1a}$ -receptor-expressing HeLa cells (Figure 5). Rather, the pretreatment of OPC-51803 at  $10^{-5}$  M inhibited the AVP-induced increase in  $[Ca^{2+}]_i$  apparently.

In HeLa cells expressing human  $V_{1b}$ -receptors, AVP ( $n=6$ ) and dDAVP ( $n=6$ ) at concentrations of  $10^{-9}$ – $10^{-6}$  M increased  $[Ca^{2+}]_i$  from  $126 \pm 9$  to  $355 \pm 17$  nM and from  $123 \pm 8$  to  $282 \pm 18$  nM, respectively. As OPC-51803 showed little affinity for  $V_{1b}$ -receptors and directly suppressed the Fura-2- $Ca^{2+}$  fluorescence at  $10^{-4}$  M, we did not evaluate the effect of OPC-51803 on intracellular  $Ca^{2+}$  concentration in HeLa cells expressing human  $V_{1b}$ -receptors.

## Discussion

In the present study, we evaluated the affinity and agonistic activity of OPC-51803, a first nonpeptide AVP  $V_2$  agonist, for human  $V_2$ -,  $V_{1a}$ -, and  $V_{1b}$ -receptors expressed in HeLa cells. OPC-51803 bound to human  $V_2$ - and  $V_{1a}$ -receptors. It increased cyclic AMP production through  $V_2$ -receptors



**Figure 5** Effect of OPC-51803, AVP, and dDAVP on  $[Ca^{2+}]_i$  in HeLa cells expressing human (a)  $V_{1a}$ - and (b)  $V_{1b}$ -receptors. Values are expressed as the mean  $\pm$  s.e. mean of five or six experiments. The experiment of OPC-51803 on agonistic activity in HeLa cells expressing  $V_{1b}$ -receptors was not performed.

though, it did not increase  $[Ca^{2+}]_i$  through V<sub>1a</sub>-receptors. On the other hand, it inhibited the AVP-induced increase in  $[Ca^{2+}]_i$  at higher concentrations. These results suggest that OPC-51803 may act rather as an antagonist for V<sub>1a</sub>-receptors than as an agonist. As to V<sub>1b</sub>-receptors, OPC-51803 inhibited only 30% of [<sup>3</sup>H]-AVP binding even at 10<sup>-4</sup> M, indicating that it has little affinity for V<sub>1b</sub>-receptors. Therefore, we did not further investigate its pharmacological properties for V<sub>1b</sub>-receptors, as it seems not to play a physiological role through V<sub>1b</sub>-receptors. These results indicate that OPC-51803 is a selective V<sub>2</sub>-receptor agonist without agonistic potencies for V<sub>1a</sub>- and V<sub>1b</sub>-receptors.

The cyclic AMP production by OPC-51803 was increased concentration-dependently in HeLa cells expressing V<sub>2</sub>-receptors. The EC<sub>50</sub> value of the cyclic AMP production is comparable to the K<sub>i</sub> value of the binding study as well as those of AVP and dDAVP. However, the maximal increase in cyclic AMP by OPC-51803 did not reach the maximal response seen with AVP or dDAVP. To confirm this partial agonism, we examined the inhibitory effect of OPC-51803 on the cyclic AMP production by AVP. OPC-51803 at concentrations of 10<sup>-9</sup>–10<sup>-5</sup> M slightly but concentration-dependently suppressed cyclic AMP production induced by a submaximal dose (10<sup>-9</sup> M) of AVP (data not shown). These results may suggest that OPC-51803 acts as a partial agonist to V<sub>2</sub>-receptors.

Among other nonpeptide agonists, it was reported that a nonpeptide angiotensin II agonist, L-162,313, showed a partial activity in *in vitro* experiments, but in *in vivo* experiments, it increased blood pressure to the same degree as angiotensin II (Kivlighn *et al.*, 1995). In preliminary experiments, we employed AVP-deficient Brattleboro rats in which diabetes insipidus is the predominant characteristic. Orally administered OPC-51803 decreased urine volume and increased urine osmolality in six animals (Hirano *et al.*, 1998). It is likely that the mechanism for this action is similar to that of dDAVP.

dDAVP is well-known as a strong and selective V<sub>2</sub>-receptor agonist and has long been utilized as a standard reagent for studies involving V<sub>2</sub>-receptors. However, since molecular cloning of human AVP receptors was achieved and the functional analysis system using their clones expressing in cells was utilized, species differences and more precise receptor selectivity of dDAVP were elucidated (Pettibone *et al.*, 1992; Saito *et al.*, 1997). dDAVP inhibited [<sup>3</sup>H]-AVP binding not only for human V<sub>2</sub>-receptors but also for V<sub>1a</sub>- and V<sub>1b</sub>-receptors, and with similar affinities. In our present study, similar results were obtained whereby the affinities of dDAVP for human AVP receptors was 13 times less potent for V<sub>1a</sub>-receptors and four times less potent for V<sub>1b</sub>-receptors compared to V<sub>2</sub>-receptors. More interestingly, dDAVP stimulated intracellular signalling through V<sub>1a</sub>- and V<sub>1b</sub>-receptors as well as through V<sub>2</sub>-receptors.

It was also recently reported that dDAVP has an agonistic activity for human V<sub>1b</sub>-receptors (Saito *et al.*, 1997); dDAVP dose-dependently stimulated inositol turnover in COS-1 cells expressing human V<sub>1b</sub>-receptors. Our results in this study are consistent with the finding that dDAVP increased intracellular Ca<sup>2+</sup> concentration in HeLa cells expressing human V<sub>1b</sub>-receptor with the similar EC<sub>50</sub> value of AVP.

As to V<sub>1a</sub>-receptors, there are some differences between their and our results. We observed a concentration-dependent increase in  $[Ca^{2+}]_i$  with an EC<sub>50</sub> of 28.9 ± 2.6 nM, while they have seen a slight increase in inositol phosphates. dDAVP-induced increase in  $[Ca^{2+}]_i$  in HeLa cells expressing human V<sub>1a</sub>-receptors was inhibited by the pretreatment of peptide V<sub>1a</sub>-antagonist (data not shown). The reason for this difference is not clear and there are few reports that dDAVP shows V<sub>1a</sub>-agonistic action for human V<sub>1a</sub>-receptors. In rats, dDAVP shows vasoconstrictor activity *via* V<sub>1a</sub>-receptors, although it is less potent than antidiuretic activity (Sawyer *et al.*, 1974a,b). It is also reported that MAP kinases were activated by dDAVP in rat 3Y1 fibroblasts through V<sub>1a</sub>-receptors (Nishioka *et al.*, 1995). These results suggest that dDAVP may have some agonistic properties for V<sub>1a</sub>-receptors.

Considering the clinical implications, dDAVP (Desmopressin<sup>TM</sup>) is currently used for the treatment of patients with central diabetes insipidus and primary nocturnal enuresis, which are AVP-deficient states. From recent studies and our results, dDAVP is not a highly selective V<sub>2</sub>-agonist; it stimulates strongly V<sub>1b</sub>-receptors and possibly V<sub>1a</sub>-receptors. In contrast, OPC-51803 does not stimulate either V<sub>1a</sub>-receptors or V<sub>1b</sub>-receptors and shows highly selective V<sub>2</sub>-receptors agonism. Although the physiological significance of these actions of dDAVP through V<sub>1a</sub>- and V<sub>1b</sub>-receptors is unclear, the specificity of OPC-51803 for V<sub>2</sub>-receptors may be desirable for the treatment of conditions involving water metabolism disorders. On the other hand, although dDAVP has antidiuretic activity when administered by the oral route, its bioavailability is less than 1% in humans and the large individual variability in the amounts or times of treatments was seen because of poor bioavailability. OPC-51803 is absorbed well and shows oral effectiveness *in vivo*. Therefore, it might offer new therapeutic advantages in the treatment of water metabolism disorders.

In conclusion, we found a novel nonpeptide human AVP V<sub>2</sub>-agonist, OPC-51803. This drug is expected to be an oral effective and practical one for patients with low circulating levels of AVP.

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